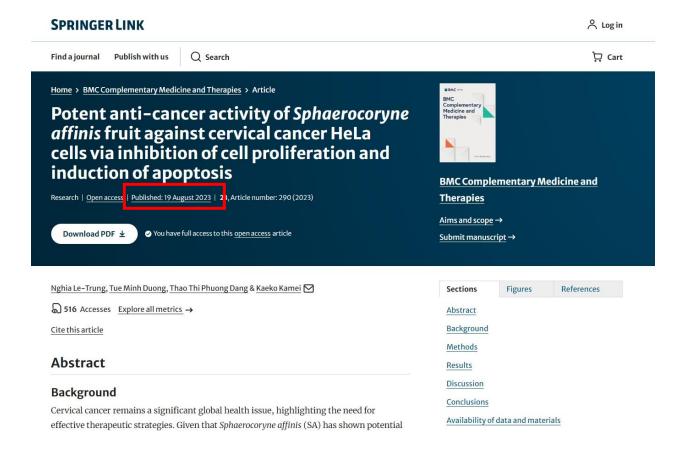
งานวิจัย เรื่อง Antiplasmodial dimeric chalcone derivatives from the roots of Uvaria siamensis มีบทความอ้างอิง 1 บทความ โดย published 19 August 20203



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#### **RESEARCH Open Access**

# Potent anti-cancer activity of Sphaerocoryne affinis fruit against cervical cancer HeLa cells via inhibition of cell proliferation and induction of apoptosis

Nghia Le-Trung<sup>1</sup>, Tue Minh Duong<sup>1</sup>, Thao Thi Phuong Dang<sup>2</sup> and Kaeko Kamei<sup>1\*</sup>

### **Abstract**

Background Cervical cancer remains a significant global health issue, highlighting the need for effective therapeutic strategies. Given that Sphaerocoryne affinis (SA) has shown potential anti-cancer activity in several cancer types, herein, we investigate the effects of SA fruit (SAF) on human cervical cancer HeLa cells and their underlying mechanisms of action.

Methods SAF extract cytotoxicity was assessed in various cancer cell lines. The effects of the hexane fraction (SAF-Hex) on HeLa cell viability, cell cycle protein expression, apoptosis, and DNA damage were evaluated using cytotoxicity assays, Western blotting, quantitative PCR, 4',6-diamidino-2-phenylindole (DAPI) staining, and a terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL) assay.

**Results** SAF-Hex selectively inhibited HeLa cell viability with an IC50 of  $4.20\pm0.36\,\mu\text{g/mL}$  and a selectivity index of 5.11 ± 0.58. The time-dependent cytotoxicity assay showed decreased cell survival after 48 h of treatment, accompanied by morphological changes and apoptotic bodies in HeLa cells. SAF-Hex also suppressed HeLa cell cycle proteins (Cyclin E, CDK2, and CDK1), reduced PCNA transcription, and diminished AKT and mTOR activation, thus inhibiting cell proliferation. The increased yH2AX expression, DNA fragmentation, and caspases-3 and -9 activation indicated SAF-Hex-induced DNA damage and apoptosis. However, the BAX/BCL-2 ratio remained unchanged, and BAX and BCL2 expression was attenuated.

Conclusion SAF-Hex effectively inhibits HeLa cell proliferation and induces DNA damage in that cervical cancer cell line activating apoptosis through the intrinsic pathway. Interestingly, the BAX/BCL-2 ratio remained unchanged while BAX and BCL2 transcription was attenuated. Hence, further research is required to explore this unexpected finding and facilitate the development of novel therapies targeting cervical cancer HeLa cells.

Keywords Anti-cancer, Apoptosis, Cervical cancer, DNA damage, HeLa cells, Sphaerocoryne affinis

Kaeko Kamei

kame@kit.ac.ip

# **Background**

Cervical cancer ranks as the fourth most common cancer and the leading cause of cancer-related deaths among women [1]. Human papillomavirus (HPV) is the primary factor in cervical cancer development, with smoking, high parity, long-term use of oral contraceptives, and sexually transmitted diseases acting as cofactors.



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<sup>\*</sup>Correspondence:

Department of Functional Chemistry, Kyoto Institute of Technology, . Kyoto 606-8585, Japan

<sup>&</sup>lt;sup>2</sup> Laboratory of Cancer Research, University of Science, Vietnam National University Ho Chi Minh City, Ho Chi Minh City, Vietnam

# **Supplementary Information**

The online version contains supplementary material available at https://doi.org/10.1186/s12906-023-04127-0.

Additional file 1: Fig. S1. Original images of blots shown in Fig. 2A. Fig. S2. Original images of blots shown in Fig. 3A. Fig. S3. Original images of blots shown in Fig. 4A. Fig. S4. Western blotting of caspase-8.

Additional file 2: Table S1. Antibodies for Western Blot Assay. Table S2. Primers for qPCR.

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#### Authors' contributions

Conceptualization: N.L., T.D.T.P., and K.K.; Investigation: N.L., D.M.T.; Methodology: N.L.; Project administration: K.K.; Supervision: K.K.; Discussion of the results: N.L., D.T.P.T., and K.K.; Writing—Original Draft: N.L.; Writing—Review and Editing: N.L., D.T.P.T., and K.K.; All authors have read and agreed to the published version of this manuscript.

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#### Availability of data and materials

The datasets generated and/or analyzed during the current study are available from the corresponding author on reasonable request.

# **Declarations**

# Ethics approval and consent to participate

The plant material used in this study, SAF (Chùm Đuông in Vietnamese), was purchased from a country market in Tay Ninh province, Vietnam. It was identified by Dr. Dang Le Anh Tuan from the Laboratory of Botany, Department of Ecology and Evolutionary Biology, Faculty of Biology—Biotechnology, University of Science, VNU-HCMC, Vietnam. A voucher specimen (PHH0004912) has been deposited and is available for reference. The collection and use of this plant material comply with the IUCN Policy Statement on Research Involving Species at Risk of Extinction and the Convention on the Trade in Endangered Species of Wild Fauna and Flora.

# Consent for publication

Not applicable.

# Competing interests

The authors declare no competing interests.

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# References

- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2021;71(3):209–49.
- Canfell K. Towards the global elimination of cervical cancer. Papillomavirus Res. 2019;8:100170.
- Johnson CA, James D, Marzan A, Armaos M. Cervical cancer: an overview of pathophysiology and management. Semin Oncol Nurs. 2019;35(2):166–74.
- Vasan N, Baselga J, Hyman DM. A view on drug resistance in cancer. Nature. 2019;575(7782):299–309.

- Zugazagoitia J, Guedes C, Ponce S, Ferrer I, Molina-Pinelo S, Paz-Ares L. Current challenges in cancer treatment. Clin Ther. 2016;38(7):1551–66.
- Jung JH, Chang CJ, Smith DL, McLaughlin JL, Pummangura S, Chaichantipyuth C, et al. Additional bioactive heptenes from *Melodorum fruticosum*. J Nat Prod. 1991;54(2):500–5.
- Tuchinda P, Udchachon J, Reutrakul V, Santisuk T, Taylor WC, Farnsworth NR, et al. Bioactive butenolides from *Melodorum fruticosum*. Phytochemistry. 1991;30(8):2685–9.
- Jung JH, Pummangura S, Chaichantipyuth C, Patarapanich C, McLaughlin JL. Bioactive constituents of Melodorum fruticosum. Phytochemistry. 1990;29(5):1667–70.
- Jung JH, Pummangura S, Chaichantipyuth C, Patarapanich C, Fanwick PE, Chang C-J, et al. New bioactive heptenes from Melodorum fruticosum (Annonaceae). Tetrahedron. 1990;46(15):5043–54.
- Chaichantipyuth C, Tiyaworanan S, Mekaroonreung S, Ngamrojnavanich N, Roengsumran S, Puthong S, et al. Oxidized heptenes from flowers of Melodorum fruticosum. Phytochemistry. 2001;58(8):1311–5.
- Nghi NBT, Uyen TT, Anh HM, Linh DM, Thao DTP. Rumdul (Sphaerocoryne affinis) antioxidant activity and its potential for Parkinson's disease treatment. Oxid Med Cell Longev. 2022;2022:8918966.
- Salae AW, Chairerk O, Sukkoet P, Chairat T, Prawat U, Tuntiwachwuttikul P, et al. Antiplasmodial dimeric chalcone derivatives from the roots of Uvaria siamensis. Phytochemistry. 2017;135:135–43.
- Amatori S, Persico G, Fanelli M. Real-time quantitative PCR array to study drug-induced changes of gene expression in tumor cell lines. J Cancer Metastasis Treat. 2017;3:90–9.
- Phan RT, Dalla-Favera R. The BCL6 proto-oncogene suppresses p53 expression in germinal-centre B cells. Nature. 2004;432(7017):635–9.
- Vermeulen K, Van Bockstaele DR, Berneman ZN. The cell cycle: a review of regulation, deregulation and therapeutic targets in cancer. Cell Prolif. 2003;36(3):131–49.
- Nitulescu GM, Van De Venter M, Nitulescu G, Ungurianu A, Juzenas P, Peng Q, et al. The AKT pathway in oncology therapy and beyond (Review). Int J Oncol. 2018;53(6):2319–31.
- Mah LJ, El-Osta A, Karagiannis TC. yH2AX: a sensitive molecular marker of DNA damage and repair. Leukemia. 2010;24(4):679–86.
- Kitazumi I, Tsukahara M. Regulation of DNA fragmentation: the role of caspases and phosphorylation. FEBS J. 2011;278(3):427–41.
- Walsh JG, Cullen SP, Sheridan C, Lüthi AU, Gerner C, Martin SJ. Executioner caspase-3 and caspase-7 are functionally distinct proteases. Proc Natl Acad Sci U S A. 2008;105(35):12815–9.
- Porter AG, Jänicke RU. Emerging roles of caspase-3 in apoptosis. Cell Death Differ. 1999;6(2):99–104.
- Yadav P, Yadav R, Jain S, Vaidya A. Caspase-3: a primary target for natural and synthetic compounds for cancer therapy. Chem Biol Drug Des. 2021;98(1):144–65.
- Hongnak S, Jongaramruong J, Khumkratok S, Siriphong P, Tip-pyang S. Chemical constituents and derivatization of melodorinol from the roots of Melodorum fruticosum. Nat Prod Commun. 2015;10(4):633–6.
- Weerapreeyakul N, Nonpunya A, Barusrux S, Thitimetharoch T, Sripanidkulchai B. Evaluation of the anticancer potential of six herbs against a hepatoma cell line. Chin Med. 2012;7(1):15.
- Indrayanto G, Putra GS, Suhud F. Chapter six validation of in vitro bioassay methods: application in herbal drug research. In: Al-Majed AA, editor. Profiles Drug Subst Excip Relat Methodol. USA: Academic Press. 2021;46:273-307.
- Saraste A, Pulkki K. Morphologic and biochemical hallmarks of apoptosis. Cardiovasc Res. 2000;45(3):528–37.
- Fagundes R, Teixeira LK. Cyclin E/CDK2: DNA replication, replication stress and genomic instability. Front Cell Dev Biol. 2021;9:774845.
- Strzalka W, Ziemienowicz A. Proliferating cell nuclear antigen (PCNA): a key factor in DNA replication and cell cycle regulation. Ann Bot. 2011;107(7):1127–40.
- Abeyrathna P, Su Y. The critical role of AKT in cardiovascular function. Vascul Pharmacol. 2015;74:38–48.
- 29. Pouliliou S, Koukourakis MI. Gamma histone 2AX (y-H2AX)as a predictive tool in radiation oncology. Biomarkers. 2014;19(3):167–80.
- Rogakou EP, Nieves-Neira W, Boon C, Pommier Y, Bonner WM. Initiation of DNA fragmentation during apoptosis induces phosphorylation of H2AX histone at serine 139. J Biol Chem. 2000;275(13):9390–5.